

R E V I E W

The vascular contribution to Alzheimer's disease

Robin ALTMAN and John C. RUTLEDGE

Department of Internal Medicine, University of California Davis, 451 East Health Sciences Drive, Davis, CA 95616, U.S.A.

A B S T R A C T

AD (Alzheimer's disease) is a progressive neurodegenerative disease of unknown origin. Despite questions as to the underlying cause(s) of this disease, shared risk factors for both AD and atherosclerotic cardiovascular disease indicate that vascular mechanisms may critically contribute to the development and progression of both AD and atherosclerosis. An increased risk of developing AD is linked to the presence of the apoE4 (apolipoprotein E4) allele, which is also strongly associated with increased risk of developing atherosclerotic cardiovascular disease. Recent studies also indicate that cardiovascular risk factors, including elevated blood cholesterol and triacylglycerol (triglyceride), increase the likelihood of AD and vascular dementia. Lipids and lipoproteins in the circulation interact intimately with the cerebrovasculature, and may have important effects on its constituent brain microvascular endothelial cells and the adjoining astrocytes, which are components of the neurovascular unit. The present review will examine the potential mechanisms for understanding the contributions of vascular factors, including lipids, lipoproteins and cerebrovascular $A\beta$ (amyloid β), to AD, and suggest therapeutic strategies for the attenuation of this devastating disease process. Specifically, we will focus on the actions of apoE, TGRLs (triacylglycerol-rich lipoproteins) and TGRL lipolysis products on injury of the neurovascular unit and increases in blood–brain barrier permeability.

INTRODUCTION

Vascular dysfunction and endothelial injury have come to be recognized as key mediators in the development of ASCVD (atherosclerotic cardiovascular disease). ASCVD is defined as a thickening of arterial walls due to accumulation of fatty material and macrophages. Atherosclerosis affects medium-large arteries and similar disease processes may also affect the smaller arteries of the brain. Although cerebrovascular disease is a major cause of mor-

bidity and mortality, research into the fundamental mechanisms underlying this problem has been relatively slow coming, compared with the emphasis that has been placed on atherosclerosis research in the last several decades.

Despite the apparent differences between the peripheral vasculature and the cerebrovasculature, many of the same mechanisms, or slight variants thereof, may also contribute to endothelial injury in the brain. Vascular dysfunction in the brain has long been recognized as a contributing factor to the development of stroke

Key words: Alzheimer's disease, apolipoprotein E (apoE), astrocyte, endothelial cell, lipoprotein, triacylglycerol-rich lipoprotein (TGRL), vascular system.

Abbreviations: ABC, ATP-binding-cassette; AD, Alzheimer's disease; APP, amyloid precursor protein; apoE, apolipoprotein E; ASCVD, atherosclerotic cardiovascular disease; $A\beta$, amyloid β ; $A\beta$ 42, $A\beta$ -(1–42); BBB, blood–brain barrier; CAA, cerebral amyloid angiopathy; CI, confidence interval; DHA, docosahexaenoic acid; HDL, high-density lipoprotein; HMG-CoA, 3-hydroxy-3-methylglutaryl-CoA; 13-HODE, (13S)-hydroxyoctadeca-(9Z,11E)-dienoic acid; HR, hazard ratio; LDL, low-density lipoprotein; LDLR, LDL receptor; LpL, lipoprotein lipase; hLpL, human LpL; LXR, liver X receptor; NEFA, non-esterified ('free') fatty acid; ROS, reactive oxygen species; SFA, saturated fatty acid; TGRL, triacylglycerol (triglyceride)-rich lipoprotein; TLR, Toll-like receptor; TNF, tumour necrosis factor; VLDL, very-low-density lipoprotein; ZO-1, zonula occludens-1.

Correspondence: Dr Robin Altman (email raltman@ucdavis.edu).

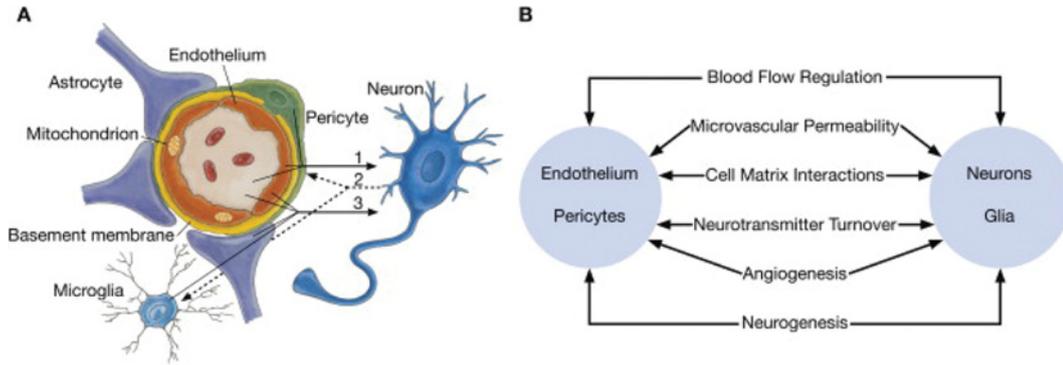


Figure 1 Schematic representation of the neurovascular unit

(A) Endothelial cells and pericytes are separated by the basement membrane. Pericyte processes sheathe most of the outer side of the basement membrane. At points of contact, pericytes communicate directly with endothelial cells through the synapse-like peg-socket contacts. Astrocytic endfoot processes unsheath the microvessel wall, which is made up of endothelial cells and pericytes. Resting microglia have a 'ramified' shape. In cases of neuronal disorders that have a primary vascular origin, circulating neurotoxins may cross the BBB to reach their neuronal targets, or pro-inflammatory signals from the vascular cells or reduced capillary blood flow may disrupt normal synaptic transmission and trigger neuronal injury (arrow 1). Microglia recruited from the blood or within the brain and the vessel wall can sense signals from neurons (arrow 2). Activated endothelium, microglia and astrocytes signal back to neurons, which in most cases aggravates the neuronal injury (arrow 3). In the case of a primary neuronal disorder, signals from neurons are sent to the vascular cells and microglia (arrow 2), which activate the vasculo-glial unit and contributes to the progression of the disease (arrow 3). (B) Co-ordinated regulation of normal neurovascular functions depends on vascular cells (endothelium and pericytes), neurons and astrocytes. Reprinted from *Neuron*, volume 57, Zlokovic, B.V., The blood-brain barrier in health and chronic neurodegenerative disorders, pp. 178–201, Copyright (2008), with permission from Elsevier (<http://www.sciencedirect.com/science/journal/08966273>).

and vascular dementia. More recently, evidence has accumulated that suggests that the development of AD (Alzheimer's disease) may also be strongly related to underlying vascular problems. Whether this vascular dysfunction actually causes AD or merely contributes to the development of the disease or worsens the disease processes once they are underway, it is worthwhile to investigate how the vasculature and its related blood lipids, lipoproteins and vascular $A\beta$ (amyloid β) promote AD. The present review will focus on the actions of apoE (apolipoprotein E), TGRLs [triacylglycerol (triglyceride)-rich lipoproteins] and TGRL lipolysis products on injury of the neurovascular unit and increases in BBB (blood-brain barrier) permeability.

PATHOPHYSIOLOGY OF AD

Dementia is the progressive decline in cognitive function due to damage or disease of the brain, and is distinct from the slowing of cognitive function that is expected with normal aging. The two most common causes of dementia are AD and vascular dementia [1–4]. For the more common form of late-onset AD, only one susceptibility allele has so far been identified, $\epsilon 4$ (E4) of the *apoE* gene [5]. In AD, progressive brain atrophy is observed, principally in the temporoparietal cortex, together with an inflammatory response of neurons and astrocytes, as well as deposition of amyloid plaques and neurofibrillary tangles. Astrocytes are a subtype of glial cells in the brain that are 'star shaped' and are key

cells in the maintenance of the BBB, as their endfeet surround endothelial cells (Figure 1) [6–11]. In addition, the many arm-like processes of astrocytes envelop neurons. Astrocytes are associated with senile plaques in the brain and inflammation of microvascular endothelial cells, and astrocytes are common features of AD [12–22].

The neurovascular unit, including brain microvascular endothelial cells and astrocytes, regulates BBB permeability. In a study examining human subjects with mild-to-moderate AD, the BBB was found to be a significant modifier of AD progression over 1 year [23]. Increased BBB permeability plays an important role in the promotion of AD by allowing potentially neurotoxic substances, such as pro-inflammatory cytokines and lipids, access to the CNS (central nervous system) [24–28].

CEREBROVASCULAR $A\beta$

The physiology and pathophysiology of the neurovascular unit in AD may well be influenced by interactions with cerebrovascular $A\beta$. $A\beta$ results from the proteolytic processing of APP (amyloid precursor protein), which is found in various cell types throughout the body, including cells of the brain. Proteolytic cleavage of APP by β -secretase and γ -secretase results in two forms of $A\beta$: $A\beta_{40}$ [$A\beta$ -(1–40)] and $A\beta_{42}$ [$A\beta$ -(1–42)] [29]. Although $A\beta$ is a physiological component of plasma, the exact origin of plasma $A\beta$ remains unknown. Peripheral sources, such as blood platelets, may prove to be important sources of plasma $A\beta$ [30]. It has been suggested that, regardless of the primary origin of plasma

$A\beta$, it may play an important role in the cerebrovascular pathology associated with AD. Higher levels of plasma $A\beta_{42}$ were found in AD patients and in those subjects who would eventually develop AD compared with those who did not develop AD [31–33]. Additionally, mutations associated with early-onset familial AD result in elevated levels of extracellular $A\beta_{42}$ [33].

The neuropathological characteristics of AD usually include sporadic CAA (cerebral amyloid angiopathy), even in the absence of underlying ASCVD, with some studies reporting up to 80% of AD patients exhibiting CAA to at least a minor degree [34,35]. Studies suggest that CAA severity increases with progressing AD [36]. The Honolulu-Asia Aging Study demonstrated that men with both CAA and AD had greater cognitive impairment than those individuals with either CAA or AD [37]. Numerous pathological, cell culture and animal model studies have demonstrated the deleterious effects of $A\beta$ peptides and CAA on cerebral microvessels [38]. This damage includes histological and ultrastructural abnormalities of cerebrovascular walls in CAA [39]. Reduced adhesion of vascular smooth muscle cells in response to treatment with $A\beta$ [40] and impaired function of vascular smooth muscle cells in transgenic mouse models of CAA [41] were also observed. Additional *in vitro* studies demonstrated that wild-type and mutant forms of $A\beta$ have anti-angiogenic and vasoactive properties [42–44]. CAA-related vascular abnormalities in both transgenic mouse models and AD patients may also contribute to capillary occlusion and altered blood flow [45,46].

The detrimental effects of plasma $A\beta$ and amyloid angiopathy on components of the cerebrovasculature suggest that cerebrovascular $A\beta$ may be intimately related to the progression and development of AD through vascular pathways. The combined influences of cerebrovascular $A\beta$, plasma lipoproteins and apoE may form a constellation of negative interactions that lead directly to vascular dysfunction. Although conflicting studies debate the usefulness of plasma $A\beta$ as a biomarker predictive of AD [32], the potential contribution of cerebrovascular $A\beta$ to the vascular pathologies associated with AD merits continued attention.

APOE STRUCTURE AND FUNCTION

Although the underlying cause of AD remains unknown, an increased risk of developing the disease is linked to the presence of the apoE4 allele. ApoE is a 34 kDa glycoprotein that has many functions, including assembly, processing and removal of plasma lipoproteins [47], neuronal repair, dendritic growth and anti-inflammatory properties [48]. It is a component apolipoprotein of VLDLs (very-low-density lipoproteins), IDLs (intermediate-density lipoproteins), chylomicrons, HDLs (high-density lipoproteins) [49] and LDLs (low-density lipoproteins) [50]. ApoE serves as a ligand for LDLRs

(LDL receptors) and, through its interaction with these receptors, participates in the distribution of cholesterol and other lipids among various cells of the body [51]. In humans, there are three common isoforms of apoE: apoE2, apoE3 and apoE4, with apoE3 being the most common isoform [52–60].

ApoE contains a 22-kDa N-terminal domain (residues 1–191) and a 10-kDa C-terminal domain (residues 222–299) connected by a protease-sensitive loop [61]. The N-terminal domain contains the LDLR-binding region (residues 136–150 in helix 4) and the C-terminal domain has a high affinity for lipid and is responsible for lipoprotein-lipid binding [62]. ApoE4 differs from apoE3 by the presence of an arginine residue at position 112, rather than a cysteine residue at the same position in apoE3 [63]. ApoE in the blood is generated by the liver, intestine and macrophages, whereas apoE in the brain is generated by glial cells, including astrocytes and microglia [64].

Among the human isoforms, apoE4 shows a unique domain interaction where the arginine residue at the 112 position induces an interaction of Arg⁶¹ in the N-terminal domain with Glu²⁵⁵ in the C-terminal domain, a feature thought to be responsible for the preferential association of apoE4 with VLDLs in blood [65,66]. Approx. 40–65% of individuals with AD have at least one copy of the E4 allele. The E4 allele is present in approx. 25% of the U.S. population [67–69] and is strongly linked to an increased risk of the development of AD and atherosclerosis complications [70–72]. ApoE4 is reported to have effects that promote amyloid deposition, neurotoxicity, oxidative stress and neurofibrillary tangle formation [48], all of which are pathophysiologically linked to AD. Clinical trials also suggest that apoE4 plays key causative roles in AD. In a postmortem study, brains from patients with advanced AD exhibited plasma proteins, such as prothrombin, in the microvessels, and suggested that increased permeability of the BBB may be more common in patients with at least one E4 allele [73]. These findings indicate that the E4 allele is an important determinate of AD.

APOE AND MODULATION OF NEURAL AND VASCULAR TISSUE RESPONSE TO INJURY

ApoE4 is associated with increased brain inflammation. Previous studies have shown that apoE4 can undergo proteolysis and cause mitochondrial damage [74], increase brain inflammation found in apoE4-expressing mice in response to LPS (lipopolysaccharide) [75] and facilitate the deposition of oligomeric $A\beta$ as amyloid to a greater extent than apoE3 [76]. Our previous study showed that apoE4 increased, and apoE3 decreased, TNF (tumour necrosis factor)- α -induced human aortic endothelial cell injury [77]. These studies indicate that, in addition to

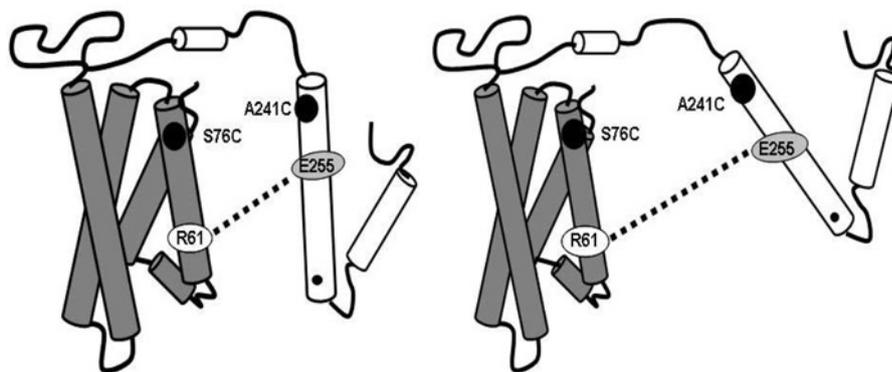


Figure 2 Schematic representation of apoE4 before and after a moderately high-fat meal

An apoE4 mutant was employed to study the domain interaction of the apoE4 mutant using EPR (electron paramagnetic resonance) spectroscopy. The dotted line represents a salt bridge between Arg⁶¹ (R61) and Glu²⁵⁵ (E255) of apoE4 showing a domain interaction. In this mutant, the serine residue at position 76 and the alanine residue at position 241 were mutated to cysteine and subsequently labelled with a nitroxyl spin label. The left-hand panel represents the structural conformation of apoE4 during the fasting state, and the right-hand panel represents the conformation in the postprandial state. ApoE4 had a reduced domain interaction during the postprandial state, implying a more linear protein [82]. The left-hand Figure was reproduced with permission from [82]. Copyright (2006) American Society for Biochemistry and Molecular Biology. The right-hand Figure was reproduced with permission from [222]. Copyright (2010) American Society for Biochemistry and Molecular Biology.

being a marker of increased risk for AD and ASCVD, apoE4 could directly generate microvascular and neural injury.

In contrast, apoE3 is associated with reduced brain inflammation [78,79]. Microglial activation by APP was reduced by apoE3 [80]. In addition, apoE3 interacts with A β via an apoE-receptor-mediated process to inhibit neurotoxicity and neuroinflammation, a process possibly related to binding and clearance of apoE3–A β oligomer complexes. Our findings suggest a role for apoE3 to prevent inflammation and balance the intracellular redox state in injured human aortic endothelial cells [77]. Thus these results suggest that apoE3 protects against and apoE4 promotes AD and ASCVD.

LIPIDS AND APOE CONFORMATION

Our studies indicate that the postprandial state, and specifically TGRL lipolysis products, can have a profound effect on apoE4 conformation to increase formation of a more linear species of apoE4 [82], which may potentially have an impact on the pathogenesis of AD (Figure 2) [82]. Regardless of the plasma triacylglycerol levels, consistent conformational changes were observed as a result of interactions between TGRL lipolysis products and apoE4. In contrast, TGRL lipolysis products did not cause linear conformation changes in apoE3. Using thrombin-accessibility assays, other studies have shown that binding of high triacylglycerol-containing VLDLs to macrophages was related to differences in the conformation of apoE [83–86], although the specific conformational change was not known. These studies, among others [72,75,82,87,88], indicate that lipids can

have dramatic effects on apoE4 conformation and binding to cells. In the presence of varying plasma lipid levels, similar apoE4 conformational and cell interaction effects were observed, which suggests that comparable mechanisms at the level of the lipoprotein may be important in apoE4 individuals irrespective of their overall lipid levels.

It is unknown whether increased VLDL particle fluidity and linearization of apoE4 resulting from VLDL lipolysis products acts to increase apoE4 binding and injury to brain microvascular endothelial cells. This potential mechanism of injury to endothelial cells may increase BBB permeability, enabling TGRL lipolysis products to have increased access to the brain, where they may promote astrocyte and neuronal injury that could initiate and perpetuate AD.

Other studies have indicated that A β clearance and degradation mechanisms in the brain are dramatically affected by the lipidation state of apoE [89–94]. Using real-time *in situ* microdialysis methods in mice, Bell et al. [95] showed that A β clearance across the BBB is decreased when A β associates with poorly lipidated apoE3, and is almost completely blocked when A β associates with lipidated apoE3. Following the same experimental methods, Deane et al. [96] showed that lipidation of the apoE2, apoE3 or apoE4 isoform significantly reduced the transport of apoE–A β complexes across the BBB. The extent of this effect was dependent on the specific isoform, where apoE4–A β transport was most significantly disrupted, compared with apoE3–A β or apoE2–A β complexes [96]. Interestingly, lipidation of apoE appears to have opposing effects on A β degradation in the brain. Additional studies have shown that strategies to increase apoE lipidation reduce amyloid burden in animal models. Overexpression

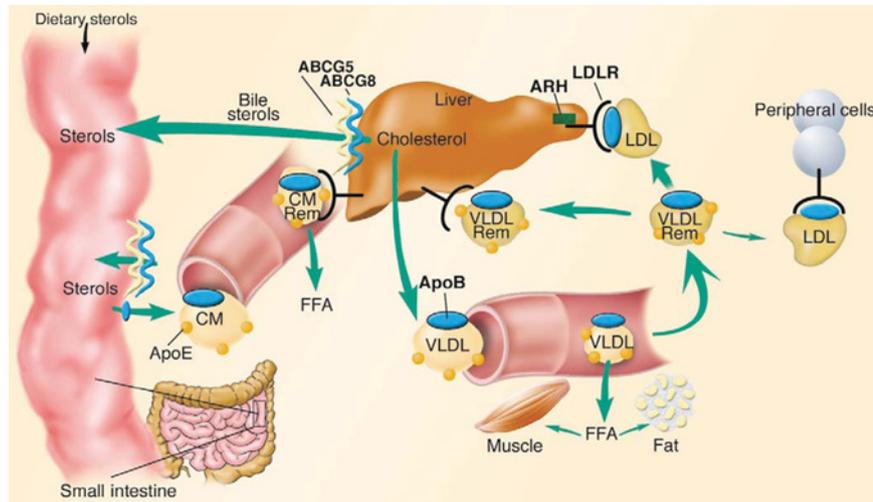


Figure 3 Overview of LDL metabolism in humans

Dietary cholesterol and triacylglycerols are packaged with apolipoproteins in the enterocytes of the small intestine and secreted into the lymphatic system as chylomicrons (CM). As chylomicrons circulate, the core triacylglycerols are hydrolysed by LpL, resulting in the formation of chylomicron remnants (CM Rem), which are rapidly removed by the liver. Dietary cholesterol has four possible fates once it reaches the liver: it can be (i) esterified and stored as cholesteryl esters in hepatocytes; (ii) packaged into VLDL particles and secreted into the plasma; (iii) secreted directly into the bile; or (iv) converted into bile acids and secreted into the bile. VLDL particles secreted into the plasma undergo lipolysis to form VLDL remnants (VLDL Rem). Approx. 50% of VLDL remnants are removed by the liver via the LDLR and the remainder mature into LDL, the major cholesterol transport particle in the blood. An estimated 70% of circulating LDL is cleared by LDLR in the liver. ABCG5 and ABCG8 (ABC family G, members 5 and 8 respectively) are located predominantly in the enterocytes of the duodenum and jejunum, the sites of uptake of dietary sterols, and in hepatocytes, where they participate in sterol trafficking into bile. ApoB, apolipoprotein B; ARH, autosomal recessive hypercholesterolaemia protein; FFA, non-esterified ('free') fatty acid. *Journal of Clinical Investigation*. Online by Rader, D.J., Cohen, J. and Hobbs, H.H. Copyright 2003 by American Society for Clinical Investigation. Reproduced with permission of American Society for Clinical Investigation in the format Journal via Copyright Clearance Center.

of ABCA1 [ABC (ATP-binding-cassette) family A1] in the brain promotes the formation of lipidated apoE and reduces the formation of amyloid plaques [97,98]. LXR (liver X receptor) agonists have also been shown to have a beneficial effect on amyloid burden in the brain. LXRs are involved with the removal of cholesterol by ABC transporters and the transfer of this cholesterol to apolipoproteins such as apoE and apoAI (apolipoprotein AI). Stimulation of these receptors by LXR agonists resulted in reduced $A\beta$ levels and enhanced apoE lipidation [92,99–101]. Furthermore, the molten globule form of apoE4, which is a partially unfolded reactive intermediate, has been associated with increased pathogenicity [74,102]. These studies indicate that the lipidation state and conformation of the apoE isoforms are important determinants of $A\beta$ homeostasis in the brain; however, the exact mechanisms by which this occurs are not well understood.

LIPOPROTEINS AND THE DEVELOPMENT OF AD

Lipoproteins are heterogeneous lipid and protein complexes. The principal function of lipoproteins is to transport lipids as fuel for cells throughout the body. Typically,

a lipoprotein particle consists of a monolayer surface shell of phospholipids, cholesterol and apolipoproteins surrounding a hydrophobic core of triacylglycerols and cholesterol esters [103–105]. The apolipoproteins are integrated into the lipid environment through amphipathic helices, which contain hydrophobic and hydrophilic domains. Following food intake, energy distribution occurs largely through generation of TGRLs, either chylomicrons or VLDLs, which are synthesized from exogenous and endogenous lipids respectively (Figure 3) [106]. The elevation of triacylglycerol in the blood after consumption of a meal, postprandial lipaemia, is the result of contributions from both of these pathways. Once in the blood, the triacylglycerols in TGRLs are hydrolysed by LpL (lipoprotein lipase), an enzyme anchored to endothelial cells [107–111]. Hydrolysis results in the successive formation of smaller TGRLs, called lipoprotein remnant particles, as well as other lipolysis products, such as fatty acids, phospholipids, monoacylglycerols and diacylglycerols. Furthermore, lipolysis of VLDL remnant particles yields LDL. The composition and size distribution of TGRLs have been shown to be important in ASCVD [56]. For example, VLDLs and chylomicron remnant lipoprotein particles are known to penetrate arterial walls and become trapped in the artery wall, thus participating in the early stages of atherosclerosis.

However, VLDL and chylomicron remnant particles are unlikely to penetrate brain microvascular endothelium because of the reduced permeability of the BBB.

Previous studies have indicated that cardiovascular risk factors, including elevated blood cholesterol and triacylglycerol, increase the likelihood of AD and dementia [112–118]. Most, but not all, previous studies have shown that elevations in mid-life cholesterol levels in the blood are associated with the development of AD [115,116,119–124]. However, definitive randomized control trials have yet to show that HMG-CoA (3-hydroxy-3-methylglutaryl-CoA) reductase inhibitors (statins) reduce the incidence of AD [59,120,125–127]. Despite the lack of conclusive evidence that cholesterol interventions have a preventative effect on AD, a substantial body of data supports the notion that mid-life blood cholesterol and triacylglycerol levels are associated with the development of AD.

TRIACYLGLYCEROLS, FATTY ACIDS AND THE INITIATION AND PROGRESSION OF AD

Most, but not all, previous studies in humans have shown an association between AD and elevated fasting triacylglycerols in the blood and also an association of AD with the metabolic syndrome. In one human study, the only significant relationship of lipids/lipoproteins with AD was elevated triacylglycerols [128]. The Three Cities Study showed that a high plasma triacylglycerol level was the only component of the metabolic syndrome that was significantly associated with the incidence of all-cause {HR (hazard ratio) 1.45, [95% CI (confidence interval), 1.05–2.00]; $P=0.02$ } and vascular [HR, 2.27 (95% CI, 1.16–4.42); $P=0.02$] dementia, even after adjustment for the apoE genotype [129]. In another study, elevated total cholesterol, LDLs and triacylglycerol, with normal HDLs and total cholesterol/HDL ratio characterize the lipid profile in AD, which overlaps with the ASCVD risk profile [130]. Additionally, patients with AD had a significantly larger mean waist circumference, higher mean plasma concentrations of triacylglycerols and glucose, and a lower mean plasma concentration of HDL-cholesterol compared with controls [131]. Thus most previous human clinical studies have shown an association of serum triacylglycerols and AD or vascular dementia.

However, the clinical measurement of the mass of triacylglycerols in blood may not reveal the true pathogenicity of TGRLs, and specifically TGRL lipolysis products. Chylomicrons and VLDL particles, which carry most triacylglycerols in blood, are not strongly associated with ASCVD or AD, perhaps because they are large particles and do not easily enter the artery wall and do not cross the BBB.

Mouse models and cell culture models also have suggested a role for triacylglycerols and fatty acids in

AD pathogenesis. Elevated triacylglycerol levels were found in the brain of a mouse model of AD [132]. In another study, elevated blood triacylglycerol levels preceded amyloid deposition in mouse brain [133]. It has been shown that fatty acids can cause generation of presenilin-1, an important determinant of γ -secretase activity necessary for generation of $A\beta$ in neuroblastoma cells [134]. Furthermore, palmitic and stearic fatty acids [SFAs (saturated fatty acids)] induce AD-like hyperphosphorylation of Tau in primary rat cortical neurons [135]. These results establish a central role for NEFAs [non-esterified ('free') fatty acids] in causing hyperphosphorylation of Tau through astroglia-mediated oxidative stress. Accumulating evidence from human, mouse and cell culture models suggests triacylglycerols and fatty acids are related to the development of AD (Figure 4) [136,137].

The literature also contains substantial data that specific fatty acids can either prevent or promote cognitive decline [119,138–141]. For example, eating fish or consuming increased long-chain omega-3 fatty acids in the diet had a beneficial effect on cognitive decline [139,142,143]. Oxidized linoleic acid, otherwise known as 13-HODE [(13*S*)-hydroxyoctadeca-(9*Z*,11*E*)-dienoic acid], is the most prevalent oxidized lipid in oxidized LDLs, a known mediator of vascular injury and atherosclerosis. A recent study has shown that 13-HODE is the most prevalent oxylipid present in VLDL lipolysis products in normal young adults [144], raising concern about life-long implications of 13-HODE elevations at the blood–endothelial cell interface where lipolysis occurs. Further study is needed to unravel the relationship between long-chain fatty acids, stroke and AD, and to examine the potential beneficial effects of omega-3 long-chain PUFAs (polyunsaturated fatty acids), such as DHA (docosahexaenoic acid) [145].

Previous studies also indicate that the fat composition and fat quantity of the diet is important in the promotion or prevention of AD, vascular dementia and ASCVD [52,53,60,146–157]. For example, a Mediterranean diet confers protection against AD, as compared with a Western diet [121,158–164]. In addition, DHA is an abundant fatty acid in the brain, and increased plasma levels of DHA were associated with reduction in AD in the Framingham Heart Study [165]. Understanding how lipids/lipoproteins influence AD pathophysiology could have a substantial impact on primary prevention of this devastating disease.

TGRL LIPOLYSIS PRODUCTS AND VASCULAR INFLAMMATION

Triacylglycerols in blood are contained in the core of TGRLs (chylomicrons, VLDLs and their remnant particles) and are not in direct contact with endothelium. LpL is anchored to brain microvascular

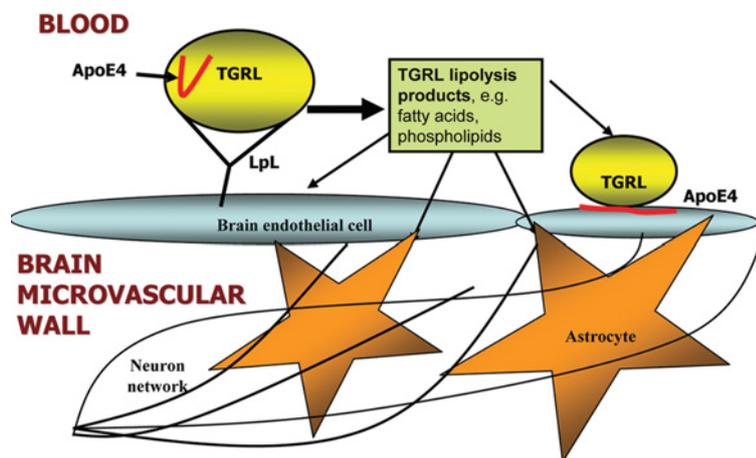


Figure 4 Vascular disease model illustrating how TGRL lipolysis products and apoE4 may interact with brain microvascular endothelial cells and astrocytes

Hydrolysis of TGRLs by LpL present in the circulation results in the release of lipolysis products, including TGRL remnant particles, mono-, di- and tri-acylglycerols, phospholipids and NEFAs. Lipolysis products may influence the inflammatory environment of the brain through two pathways: (i) direct injury to brain microvascular endothelial cells or astrocytes, or (ii) indirect injury to glial cells and neurons through cascades that begin with damaged endothelial cells. In addition, apoE4 associated with TGRL undergoes a conformational change to a more linear species in the presence of TGRL lipolysis products. This conformational change may influence the binding of apoE4 to brain microvascular endothelial cells. Negative effects to the endothelial cells due to the altered apoE4 conformation may disrupt the barrier function of the cerebrovasculature and allow TGRL lipolysis products to access the brain parenchyma, causing damage to neurons and glial cells.

endothelium, where it hydrolyses TGRLs to smaller lipolysis products, such as fatty acids and phospholipids. These lipolysis products are generated in very high concentrations immediately adjacent to brain microvascular endothelial cells. Potentially, lipolysis products generated at the luminal surface of the vascular endothelium can directly injure the endothelium and increase permeability and/or cross the BBB and injure astrocytes (Figure 4). Our studies have shown that TGRL particles, including chylomicrons and VLDLs, have relatively little effect on a variety of endothelial cell types in comparison with the dramatic effects on endothelial cell injury that we have observed with TGRL lipolysis products [107,166–169]. Thus, with regard to the brain microvasculature, TGRL lipolysis products, rather than plasma triacylglycerols, may be the most meaningful lipids to study in terms of the pathogenesis of AD.

Previous investigations have shown that TGRL lipolysis products have both pro- and anti-inflammatory effects on endothelium [170–173]. Sub-physiological concentrations of lipolysis products (5–10 μg of VLDL + 200 units/ml LpL) in endothelial cell culture prevented endothelial cell injury through generation of PPAR (peroxisome-proliferator-activated receptor) ligands in response to cytokine stimulation with TNF- α [174,175]. Similar studies using human aortic endothelial cells in culture generated comparable results. However, as VLDL triacylglycerol concentrations approached physiological concentrations (50 mg/dl triacylglycerol) in the human aortic endothelial cell culture system,

endothelial cell injury predominated when TGRLs were treated with LpL. Thus TGRL lipolysis products at high physiological-to-pathophysiological concentrations appear to have a predominately pro-inflammatory effect.

The products of TGRL lipolysis, especially the fatty acid fraction, and treatment with individual fatty acids, such as palmitic acid, in moderate-to-high physiological concentrations also injure endothelial cells. These experiments strongly indicate that it is the enzymatic function of LpL that is an important factor in generating endothelial cell injury [166]. As shown by Eiselein et al. [166] in Figure 5, the permeability of human aortic endothelial cell monolayers in culture is radically affected by exposure to TGRL lipolysis products. ZO-1 (zonula occludens-1) is an integral member of structurally sound tight junctions between adjacent endothelial cells and helps to regulate the paracellular permeability of endothelial monolayers. Exposure to TGRL lipolysis products resulted in a transition from smooth continuous ZO-1 staining to a fragmented discontinuous appearance. The disruption of the structural integrity of the tight junctions between cells suggests that the monolayer's ability to regulate its permeability was negatively affected. Additional experiments using TEER (transendothelial electrical resistance) measurements showed that the resistance of the monolayer decreased following treatment with TGRL lipolysis products [166]. Recent studies in mice overexpressing hLpL (human LpL) have shown that excess vascular wall LpL augments vascular dysfunction

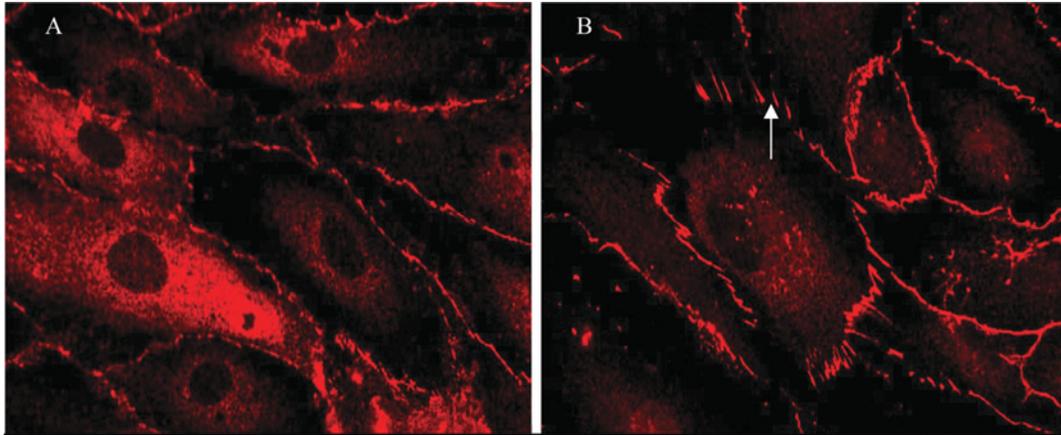


Figure 5 ZO-1 immunofluorescence before and after treatment with TGRL lipolysis products

Images of human aortic endothelial cell monolayers showing (A) continuous ZO-1 staining with treatment with TGRL (150 mg/dl triacylglycerol), and (B) discontinuity and radial rearrangement of ZO-1 (arrow) during exposure to lipolysis products generated from co-incubation of TGRL (150 mg/dl) + LpL (2 units/ml), causing increased endothelial layer permeability. This Figure was reproduced from [166] and is used with permission. Copyright (2007) American Physiological Society.

in the setting of inflammation [176,177]. Furthermore, in transgenic mice expressing hLpL, agonist-induced contraction of smooth muscle cells was increased when compared with that of wild-type mice [178]. These studies strongly indicate that TGRL lipolysis products in high-physiological and supra-physiological concentrations injure endothelium and smooth muscle cells. The aforementioned experiments that observed similar results using varying concentrations of LpL in both cell culture and animal models suggest that the physiological action of normal LpL levels is sufficient to generate injurious levels of TGRL lipolysis products at the vascular wall.

Lipotoxicity is the term commonly used to describe cell dysfunction and death induced by lipid accumulation in non-adipose tissue [179]. Most lipotoxicity has been associated with SFA-induced lipotoxicity [180–182]. Furthermore, lipotoxicity has been related to ER (endoplasmic reticulum) stress [181,183,184], apoptosis [182], mitochondrial dysfunction [185–188], lysosomal pathways and potentially autophagy [189,190]. Neurovascular lipotoxicity may play an important role in TGRL lipolysis-induced brain microvascular endothelial cell and astrocyte injury.

TGRL LIPOLYSIS PRODUCTS, ROS (REACTIVE OXYGEN SPECIES) AND AD

Oxidative stress, in which ROS overwhelm antioxidant mechanisms, is hypothesized to be important in neurodegeneration [191–193]. Previous studies have shown ROS to be important in AD [2,12,14,21,194–197], although whether ROS are causes or consequences of AD has yet to be defined. For example, lipoprotein oxidizability (as indicated by the accumulation of lipid hydroperox-

ides from the aggregate lipoproteins under oxidizing conditions) was measured in cerebrospinal fluid and plasma from 29 AD patients and was found to be significantly increased in comparison with 29 controls without dementia [198]. ROS can directly oxidize and damage DNA, proteins and lipids [199], and induce stress-response genes. In addition, astrocyte NADPH oxidase may be a key enzyme in the production of ROS in AD [12]. Some studies have indicated that ROS can be reduced by HMG-CoA reductase inhibitors (statins) [88]. One potential mechanism by which statins could reduce astrocyte injury is by down-regulation of NADPH oxidase, leading to a reduction in ROS production [200,201]. In addition, ROS can mediate apoptosis by mitochondrial apoptotic pathways [202]. VLDL lipolysis products generate ROS in human aortic endothelial cells [144,203], suggesting that VLDL lipolysis product-induced ROS generation in human microvascular endothelial cells and astrocytes may be an important mechanism of oxidative injury in these cells.

TLRs (Toll-like receptors) are a class of proteins that play a key role in the innate immune system. Recent work has shown that TLRs may be important in the pathophysiology of AD and may specifically mediate astrocyte injury and apoptosis [204,205]. SFAs modulate TLR4 through regulation of receptor dimerization and incorporation of TLR4 into lipid rafts in a ROS-dependent manner [206]. High concentrations of SFAs induced by lipolysis of VLDLs may injure astrocytes by generation of ROS and activation of TLR4.

BRAIN LIPIDS AND AD

The brain contains more lipid than any other single organ in the body. Both extra- and intra-cellular lipids,

as well as lipids within the plasma membrane, are essential to normal brain function, but also have the capacity to induce injury, apoptosis and cell death of the brain [207–218]. DHA, phospholipid oxidation products, neurotrophin receptors, lipoprotein receptors, neural membrane glycerophospholipid and sphingolipid mediators, and small-molecule oxidation products that trigger disease-associated protein misfolding have all been associated with either the promotion or prevention of AD. Montine et al. [219] demonstrated that measurements of F₂-isoprostanes in cerebrospinal fluid, which are indicative of free radical damage to brain lipids, correlate well with the degree of neurodegeneration in AD patients and may be a promising biomarker for AD. Preliminary studies also suggest that resolvins, compounds that are derived from the omega-3 fatty acids EPA (eicosapentaenoic acid) and DHA [220,221], may be important in AD for their anti-inflammatory properties. Further research is needed to unravel the intricate web of connections between specific lipids, their locations and functions within the brain, and the pathology of AD.

CONCLUSIONS AND CLINICAL IMPLICATIONS

Current treatment strategies for AD revolve around attempts to slow the progression of the disease, maintain cognitive ability and reduce the negative behavioural consequences of the disease. However, there are no known treatments to prevent or cure AD. As the underlying causative factors for AD are still unknown, targeted treatments that effectively combat the development of the disease have proven to be elusive. An increasing number of studies suggest that, whether or not lifestyle factors are the primary cause of AD, they have a significant effect on the course of the disease. **As such, these lifestyle factors represent an important avenue of approach to help slow or prevent the progression of AD. In particular, lipids derived from the diet, such as fatty acids, provide an appealing target, given the abundance of current research that suggests potentially detrimental effects of these lipids on cellular components of the cerebrovasculature.**

Our present review shows considerable overlap of the risk factors associated with AD and the risk factors associated with ASCVD, such as dyslipidaemia and the presence of the apoE4 allele. Furthermore, dyslipidaemia, hypertension and diabetes are believed to be causative for ASCVD. It is reasonable to speculate that these same risk factors could initiate or promote AD, although these areas are relatively understudied in AD. Given this background information, aggressive treatment of 'cardiovascular risk factors' for the prevention and attenuation of AD appears warranted and should be vigorously pursued.

FUNDING

R.A. was supported by an American Heart Association Western States Affiliate Predoctoral Fellowship. J.R. was supported by a National Institutes of Health (NIH) Research Program Grant (R01) [grant number HL55667], a UC Davis AD Center Core Pilot and Feasibility Project [grant number P30 AG10129-15], and the Richard A. and Nora Eccles Harrison Endowed Chair in Diabetes Research. This publication was made possible by grant number UL1 RR024146 to the UC Davis Clinical and Translational Science Center from the National Center for Research Resources (NCR), a component of NIH and NIH Roadmap for Medical Research. Its contents are solely the responsibility of the authors and do not necessarily represent the official view of NCR or NIH. Information on Re-engineering the Clinical Research Enterprise can be obtained from <http://nihroadmap.nih.gov/clinicalresearch/overview-translational.asp>.

REFERENCES

- 1 Aggarwal, N. T. and Decarli, C. (2007) Vascular dementia: emerging trends. *Semin. Neurol.* **27**, 66–77
- 2 Chui, H. C., Zarow, C., Mack, W. J., Ellis, W. G., Zheng, L., Jagust, W. J., Mungas, D., Reed, B. R., Kramer, J. H., Decarli, C. C. et al. (2006) Cognitive impact of subcortical vascular and Alzheimer's disease pathology. *Ann. Neurol.* **60**, 677–687
- 3 DeCarli, C. (2003) The role of cerebrovascular disease in dementia. *Neurologist* **9**, 123–136
- 4 DeCarli, C. S. (2006) When two are worse than one: stroke and Alzheimer disease. *Neurology* **67**, 1326–1327
- 5 Berglund, L., Wiklund, O., Eggertsen, G., Olofsson, S. O., Eriksson, M., Linden, T., Bondjers, G. and Angelin, B. (1993) Apolipoprotein E phenotypes in familial hypercholesterolaemia: importance for expression of disease and response to therapy. *J. Intern. Med.* **233**, 173–178
- 6 Abbott, N. J., Ronnback, L. and Hansson, E. (2006) Astrocyte-endothelial interactions at the blood-brain barrier. *Nat. Rev. Neurosci.* **7**, 41–53
- 7 Benarroch, E. E. (2007) Neurovascular unit dysfunction: a vascular component of Alzheimer disease? *Neurology* **68**, 1730–1732
- 8 Gee, J. R. and Keller, J. N. (2005) Astrocytes: regulation of brain homeostasis via apolipoprotein E. *Int. J. Biochem. Cell Biol.* **37**, 1145–1150
- 9 Haseloff, R. F., Blasig, I. E., Bauer, H. C. and Bauer, H. (2005) In search of the astrocytic factor(s) modulating blood-brain barrier functions in brain capillary endothelial cells *in vitro*. *Cell. Mol. Neurobiol.* **25**, 25–39
- 10 Parri, R. and Crunelli, V. (2003) An astrocyte bridge from synapse to blood flow. *Nat. Neurosci.* **6**, 5–6
- 11 Žlokovic, B. V. (2008) The blood-brain barrier in health and chronic neurodegenerative disorders. *Neuron* **57**, 178–201
- 12 Abramov, A. Y. and Duchen, M. R. (2005) The role of an astrocytic NADPH oxidase in the neurotoxicity of amyloid β peptides. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **360**, 2309–2314
- 13 Blasko, I., Stampfer-Kountchev, M., Robatscher, P., Veerhuis, R., Eikelenboom, P. and Grubeck-Loebenstien, B. (2004) How chronic inflammation can affect the brain and support the development of Alzheimer's disease in old age: the role of microglia and astrocytes. *Aging Cell* **3**, 169–176

- 14 Drew, P. D., Xu, J., Storer, P. D., Chavis, J. A. and Racke, M. K. (2006) Peroxisome proliferator-activated receptor agonist regulation of glial activation: relevance to CNS inflammatory disorders. *Neurochem. Int.* **49**, 183–189
- 15 Huang, Y., Weisgraber, K. H., Mucke, L. and Mahley, R. W. (2004) Apolipoprotein E: diversity of cellular origins, structural and biophysical properties, and effects in Alzheimer's disease. *J. Mol. Neurosci.* **23**, 189–204
- 16 Husemann, J. and Silverstein, S. C. (2001) Expression of scavenger receptor class B, type I, by astrocytes and vascular smooth muscle cells in normal adult mouse and human brain and in Alzheimer's disease brain. *Am. J. Pathol.* **158**, 825–832
- 17 Kadiu, I., Glanzer, J. G., Kipnis, J., Gendelman, H. E. and Thomas, M. P. (2005) Mononuclear phagocytes in the pathogenesis of neurodegenerative diseases. *Neurotox. Res.* **8**, 25–50
- 18 Miao, H., Hu, Y. L., Shiu, Y. T., Yuan, S., Zhao, Y., Kaunas, R., Wang, Y., Jin, G., Usami, S. and Chien, S. (2005) Effects of flow patterns on the localization and expression of VE-cadherin at vascular endothelial cell junctions: in vivo and *in vitro* investigations. *J. Vasc. Res.* **42**, 77–89
- 19 Minagar, A., Shapshak, P., Fujimura, R., Ownby, R., Heyes, M. and Eisdorfer, C. (2002) The role of macrophage/microglia and astrocytes in the pathogenesis of three neurologic disorders: HIV-associated dementia, Alzheimer disease, and multiple sclerosis. *J. Neurol. Sci.* **202**, 13–23
- 20 Rossner, S. (2004) New players in old amyloid precursor protein-processing pathways. *Int. J. Dev. Neurosci.* **22**, 467–474
- 21 Sastre, M., Klockgether, T. and Heneka, M. T. (2006) Contribution of inflammatory processes to Alzheimer's disease: molecular mechanisms. *Int. J. Dev. Neurosci.* **24**, 167–176
- 22 Strazielle, N., Gherzi-Egea, J. F., Ghiso, J., Dehouck, M. P., Frangione, B., Patlak, C., Fenstermacher, J. and Gorevic, P. (2000) *In vitro* evidence that β -amyloid peptide 1–40 diffuses across the blood-brain barrier and affects its permeability. *J. Neuropathol. Exp. Neurol.* **59**, 29–38
- 23 Bowman, G. L., Kaye, J. A., Moore, M., Waichunas, D., Carlson, N. E. and Quinn, J. F. (2007) Blood-brain barrier impairment in Alzheimer disease: stability and functional significance. *Neurology* **68**, 1809–1814
- 24 Bell, R. D. and Zlokovic, B. V. (2009) Neurovascular mechanisms and blood-brain barrier disorder in Alzheimer's disease. *Acta Neuropathol.* **118**, 103–113
- 25 Desai, T. R., Leeper, N. J., Hynes, K. L. and Gewertz, B. L. (2002) Interleukin-6 causes endothelial barrier dysfunction via the protein kinase C pathway. *J. Surg. Res.* **104**, 118–123
- 26 Lashuel, H. A. (2005) Membrane permeabilization: a common mechanism in protein-misfolding diseases. *Sci. Aging Knowl. Environ.* **2005**, pe28
- 27 Persidsky, Y., Ramirez, S., Haorah, J. and Kanmogne, G. (2006) Blood-brain barrier: structural components and function under physiologic and pathologic conditions. *J. Neuroimmune Pharmacol.* **1**, 223–236
- 28 Simionescu, M. and Antohe, F. (2006) Functional ultrastructure of the vascular endothelium: changes in various pathologies. *Handb. Exp. Pharmacol.* **176**, 41–69
- 29 Saido, T. C. and Iwata, N. (2006) Metabolism of amyloid β peptide and pathogenesis of Alzheimer's disease: towards presymptomatic diagnosis, prevention and therapy. *Neurosci. Res.* **54**, 235–253
- 30 Skovronsky, D. M., Lee, V. M. and Pratico, D. (2001) Amyloid precursor protein and amyloid β peptide in human platelets. Role of cyclooxygenase and protein kinase C. *J. Biol. Chem.* **276**, 17036–17043
- 31 Mayeux, R., Tang, M. X., Jacobs, D. M., Manly, J., Bell, K., Merchant, C., Small, S. A., Stern, Y., Wisniewski, H. M. and Mehta, P. D. (1999) Plasma amyloid β -peptide 1–42 and incipient Alzheimer's disease. *Ann. Neurol.* **46**, 412–416
- 32 Mayeux, R. M., Honig, L. S. M., Tang, M. X. P., Manly, J. P., Stern, Y. P., Schupf, N. P. and Mehta, P. D. P. (2003) Plasma A β 40 and A β 42 and Alzheimer's disease: relation to age, mortality, and risk. *Neurology* **61**, 1185–1190
- 33 Scheuner, D., Eckman, C., Jensen, M., Song, X., Citron, M., Suzuki, N., Bird, T. D., Hardy, J., Hutton, M., Kukull, W. et al. (1996) Secreted amyloid β -protein similar to that in the senile plaques of Alzheimer's disease is increased in vivo by the presenilin 1 and 2 and APP mutations linked to familial Alzheimer's disease. *Nat. Med.* **2**, 864–870
- 34 Ellis, R. J., Olichney, J. M., Thal, L. J., Mirra, S. S., Morris, J. C., Beekly, D. and Heyman, A. (1996) Cerebral amyloid angiopathy in the brains of patients with Alzheimer's disease: the CERAD experience, Part XV. *Neurology* **46**, 1592–1596
- 35 Attems, J., Quass, M., Jellinger, K. A. and Lintner, F. (2007) Topographical distribution of cerebral amyloid angiopathy and its effect on cognitive decline are influenced by Alzheimer disease pathology. *J. Neurol. Sci.* **257**, 49–55
- 36 Attems, J., Jellinger, K. A. and Lintner, F. (2005) Alzheimer's disease pathology influences severity and topographical distribution of cerebral amyloid angiopathy. *Acta Neuropathol.* **110**, 222–231
- 37 Pfeifer, L. A. M., White, L. R. M., Ross, G. W. M., Petrovitch, H. M. and Launer, L. J. P. (2002) Cerebral amyloid angiopathy and cognitive function: the HAAS autopsy study. *Neurology* **58**, 1629–1634
- 38 Herzig, M. C., Van Nostrand, W. E. and Jucker, M. (2006) Mechanism of cerebral β -amyloid angiopathy: murine and cellular models. *Brain Pathol.* **16**, 40–54
- 39 Vinters, H. V., Secor, D. L., Read, S. L., Frazee, J. G., Tomiyasu, U., Stanley, T. M., Ferreiro, J. A. and Akers, M. A. (1994) Microvasculature in brain biopsy specimens from patients with Alzheimer's disease: an immunohistochemical and ultrastructural study. *Ultrastruct. Pathol.* **18**, 333–348
- 40 Mok, S. S., Losic, D., Barrow, C. J., Turner, B. J., Masters, C. L., Martin, L. L. and Small, D. H. (2006) The β -amyloid peptide of Alzheimer's disease decreases adhesion of vascular smooth muscle cells to the basement membrane. *J. Neurochem.* **96**, 53–64
- 41 Christie, R., Yamada, M., Moskowitz, M. and Hyman, B. (2001) Structural and functional disruption of vascular smooth muscle cells in a transgenic mouse model of amyloid angiopathy. *Am. J. Pathol.* **158**, 1065–1071
- 42 Paris, D., Ait-Ghezala, G., Mathura, V. S., Patel, N., Quadros, A., Laporte, V. and Mullan, M. (2005) Anti-angiogenic activity of the mutant Dutch A β peptide on human brain microvascular endothelial cells. *Brain Res. Mol. Brain Res.* **136**, 212–230
- 43 Paris, D., Humphrey, J., Quadros, A., Patel, N., Crescentini, R., Crawford, F. and Mullan, M. (2003) Vasoactive effects of A β in isolated human cerebrovessels and in a transgenic mouse model of Alzheimer's disease: role of inflammation. *Neurol. Res.* **25**, 642–651
- 44 Paris, D., Townsend, K., Quadros, A., Humphrey, J., Sun, J., Brem, S., Wotoczek-Obadia, M., DelleDonne, A., Patel, N., Obregon, D. F. et al. (2004) Inhibition of angiogenesis by A β peptides. *Angiogenesis* **7**, 75–85
- 45 Stopa, E. G. M. D., Butala, P. M. D., Salloway, S. M. D., Johanson, C. E. P., Gonzalez, L. P., Tavares, R. B., Hovanesian, V. B., Hulette, C. M. M. D., Vitek, M. P. P. and Cohen, R. A. P. (2008) Cerebral cortical arteriolar angiopathy, vascular β -amyloid, smooth muscle actin, Braak stage, and ApoE genotype. *Stroke* **39**, 814–821
- 46 Thal, D. R., Capetillo-Zarate, E., Larionov, S., Staufenbiel, M., Zurbuegg, S. and Beckmann, N. (2009) Capillary cerebral amyloid angiopathy is associated with vessel occlusion and cerebral blood flow disturbances. *Neurobiol. Aging* **30**, 1936–1948
- 47 Fazio, S., Linton, M. F. and Swift, L. L. (2000) The cell biology and physiologic relevance of ApoE recycling. *Trends Cardiovasc. Med.* **10**, 23–30
- 48 Lahiri, D. K. (2004) Apolipoprotein E as a target for developing new therapeutics for Alzheimer's disease based on studies from protein, RNA, and regulatory region of the gene. *J. Mol. Neurosci.* **23**, 225–233

- 49 Miserez, A. R., Scharnagl, H., Muller, P. Y., Mirsaidi, R., Stahelin, H. B., Monsch, A., Marz, W. and Hoffmann, M. M. (2003) Apolipoprotein E3Basel: new insights into a highly conserved protein region. *Eur. J. Clin. Invest.* **33**, 677–685
- 50 Karlsson, H., Leanderson, P., Tagesson, C. and Lindahl, M. (2005) Lipoproteomics I: mapping of proteins in low-density lipoprotein E2 on coronary artery disease in African Americans is mediated through lipoprotein cholesterol. *J. Lipid Res.* **47**, 2475–2481
- 51 Mahley, R. W. (1988) Apolipoprotein E: cholesterol transport protein with expanding role in cell biology. *Science* **240**, 622–630
- 52 Anuurad, E., Rubin, J., Lu, G., Pearson, T. A., Holleran, S., Ramakrishnan, R. and Berglund, L. (2006) Protective effect of apolipoprotein E2 on coronary artery disease in African Americans is mediated through lipoprotein cholesterol. *J. Lipid Res.* **47**, 2475–2481
- 53 Berglund, L. (2001) The APOE gene and diets–food (and drink) for thought. *Am. J. Clin. Nutr.* **73**, 669–670
- 54 Eggertsen, G., Heimburger, O., Stenvinkel, P. and Berglund, L. (1997) Influence of variation at the apolipoprotein E locus on lipid and lipoprotein levels in CAPD patients. *Nephrol. Dial. Transplant.* **12**, 141–144
- 55 Eggertsen, G., Tegelman, R., Ericsson, S., Angelin, B. and Berglund, L. (1993) Apolipoprotein E polymorphism in a healthy Swedish population: variation of allele frequency with age and relation to serum lipid concentrations. *Clin. Chem.* **39**, 2125–2129
- 56 Hyson, D., Rutledge, J. C. and Berglund, L. (2003) Postprandial lipemia and cardiovascular disease. *Curr. Atheroscler. Rep.* **5**, 437–444
- 57 Isasi, C. R., Shea, S., Deckelbaum, R. J., Couch, S. C., Starc, T. J., Otvos, J. D. and Berglund, L. (2000) Apolipoprotein $\epsilon 2$ allele is associated with an anti-atherogenic lipoprotein profile in children: the Columbia University BioMarkers Study. *Pediatrics* **106**, 568–575
- 58 Pablos-Mendez, A., Mayeux, R., Ngai, C., Shea, S. and Berglund, L. (1997) Association of apo E polymorphism with plasma lipid levels in a multiethnic elderly population. *Arterioscler. Thromb. Vasc. Biol.* **17**, 3534–3541
- 59 Romas, S. N., Tang, M. X., Berglund, L. and Mayeux, R. (1999) APOE genotype, plasma lipids, lipoproteins, and AD in community elderly. *Neurology* **53**, 517–521
- 60 Rubin, J. and Berglund, L. (2002) Apolipoprotein E and diets: a case of gene–nutrient interaction? *Curr. Opin. Lipidol.* **13**, 25–32
- 61 Wetterau, J. R., Aggerbeck, L. P., Rall, Jr, S. C. and Weisgraber, K. H. (1988) Human apolipoprotein E3 in aqueous solution. I. Evidence for two structural domains. *J. Biol. Chem.* **263**, 6240–6248
- 62 Wilson, C., Wardell, M. R., Weisgraber, K. H., Mahley, R. W. and Agard, D. A. (1991) Three-dimensional structure of the LDL receptor-binding domain of human apolipoprotein E. *Science* **252**, 1817–1822
- 63 Weisgraber, K. H., Rall, Jr, S. C. and Mahley, R. W. (1981) Human E apoprotein heterogeneity. Cysteine–arginine interchanges in the amino acid sequence of the apo-E isoforms. *J. Biol. Chem.* **256**, 9077–9083
- 64 Vance, J. E., Karten, B. and Hayashi, H. (2006) Lipid dynamics in neurons. *Biochem. Soc. Trans.* **34**, 399–403
- 65 Dong, L. M. and Weisgraber, K. H. (1996) Human apolipoprotein E4 domain interaction. Arginine 61 and glutamic acid 255 interact to direct the preference for very low density lipoproteins. *J. Biol. Chem.* **271**, 19053–19057
- 66 Saito, H., Dhanasekaran, P., Baldwin, F., Weisgraber, K. H., Phillips, M. C. and Lund-Katz, S. (2003) Effects of polymorphism on the lipid interaction of human apolipoprotein E. *J. Biol. Chem.* **278**, 40723–40729
- 67 Dallongeville, J., Davignon, J. and Lussier-Cacan, S. (1992) ACAT activity in freshly isolated human mononuclear cell homogenates from hyperlipidemic subjects. *Metab. Clin. Exp.* **41**, 154–159
- 68 Fleisher, A., Grundman, M., Jack, Jr, C. R., Petersen, R. C., Taylor, C., Kim, H. T., Schiller, D. H., Bagwell, V., Sencakova, D., Weiner, M. F. et al. (2005) Sex, apolipoprotein E ϵ 4 status, and hippocampal volume in mild cognitive impairment. *Arch. Neurol.* **62**, 953–957
- 69 Tiret, L., de Knijff, P., Menzel, H. J., Ehnholm, C., Nicaud, V. and Havekes, L. M. (1994) ApoE polymorphism and predisposition to coronary heart disease in youths of different European populations. The EARS Study. European Atherosclerosis Research Study. *Arterioscler. Thromb.* **14**, 1617–1624
- 70 Davignon, J., Gregg, R. E. and Sing, C. F. (1988) Apolipoprotein E polymorphism and atherosclerosis. *Arteriosclerosis* **8**, 1–21
- 71 Roses, A. D., Einstein, G., Gilbert, J., Goedert, M., Han, S. H., Huang, D., Hulette, C., Masliah, E., Pericak-Vance, M. A., Saunders, A. M. et al. (1996) Morphological, biochemical, and genetic support for an apolipoprotein E effect on microtubular metabolism. *Ann. N.Y. Acad. Sci.* **777**, 146–157
- 72 Weisgraber, K. H. and Mahley, R. W. (1996) Human apolipoprotein E: the Alzheimer's disease connection. *FASEB J.* **10**, 1485–1494
- 73 Zipser, B. D., Johanson, C. E., Gonzalez, L., Berzin, T. M., Tavares, R., Hulette, C. M., Vitek, M. P., Hovanessian, V. and Stopa, E. G. (2007) Microvascular injury and blood-brain barrier leakage in Alzheimer's disease. *Neurobiol. Aging* **28**, 977–986
- 74 Mahley, R. W., Weisgraber, K. H. and Huang, Y. (2006) Apolipoprotein E4: a causative factor and therapeutic target in neuropathology, including Alzheimer's disease. *Proc. Natl. Acad. Sci. U.S.A.* **103**, 5644–5651
- 75 Ophir, G., Amariglio, N., Jacob-Hirsch, J., Elkou, R., Rechavi, G. and Michaelson, D. M. (2005) Apolipoprotein E4 enhances brain inflammation by modulation of the NF- κ B signaling cascade. *Neurobiol. Dis.* **20**, 709–718
- 76 Manelli, A. M., Stine, W. B., Van Eldik, L. J. and LaDu, M. J. (2004) ApoE and A β 1–Abet42 interactions: effects of isoform and conformation on structure and function. *J. Mol. Neurosci.* **23**, 235–246
- 77 Mullick, A. E., Powers, A. F., Kota, R. S., Tetali, S. D., Eiserich, J. P. and Rutledge, J. C. (2007) Apolipoprotein E3- and nitric oxide-dependent modulation of endothelial cell inflammatory responses. *Arterioscler. Thromb. Vasc. Biol.* **27**, 339–345
- 78 Ophir, G., Meilin, S., Efrati, M., Chapman, J., Karussis, D., Roses, A. and Michaelson, D. M. (2003) Human apoE3 but not apoE4 rescues impaired astrocyte activation in apoE null mice. *Neurobiol. Dis.* **12**, 56–64
- 79 Rubinsztein, D. C., Hanlon, C. S., Irving, R. M., Goodburn, S., Evans, D. G., Kellar-Wood, H., Xuereb, J. H., Bandmann, O. and Harding, A. E. (1994) Apo E genotypes in multiple sclerosis, Parkinson's disease, schwannomas and late-onset Alzheimer's disease. *Mol. Cell. Probes* **8**, 519–525
- 80 Barger, S. W. and Harmon, A. D. (1997) Microglial activation by Alzheimer amyloid precursor protein and modulation by apolipoprotein E. *Nature* **388**, 878–881
- 81 Reference deleted
- 82 Tetali, S. D., Budamagunta, M. S., Voss, J. C. and Rutledge, J. C. (2006) C-terminal interactions of apolipoprotein E4 respond to the postprandial state. *J. Lipid Res.* **47**, 1358–1365
- 83 Bradley, W. A. and Gianturco, S. H. (1986) ApoE is necessary and sufficient for the binding of large triglyceride-rich lipoproteins to the LDL receptor; apoB is unnecessary. *J. Lipid Res.* **27**, 40–48
- 84 Brown, S. A., Via, D. P., Gotto, Jr, A. M., Bradley, W. A. and Gianturco, S. H. (1986) Apolipoprotein E-mediated binding of hypertriglyceridemic very low density lipoproteins to isolated low density lipoprotein receptors detected by ligand blotting. *Biochem. Biophys. Res. Commun.* **139**, 333–340

- 85 Gianturco, S. H. and Bradley, W. A. (1986) Interactions of triglyceride-rich lipoproteins with receptors: modulation by thrombin. *Semin. Thromb. Hemostasis* **12**, 277–279
- 86 Gianturco, S. H., Gotto, Jr, A. M., Hwang, S. L., Karlin, J. B., Lin, A. H., Prasad, S. C. and Bradley, W. A. (1983) Apolipoprotein E mediates uptake of Sf 100–400 hypertriglyceridemic very low density lipoproteins by the low density lipoprotein receptor pathway in normal human fibroblasts. *J. Biol. Chem.* **258**, 4526–4533
- 87 Hatters, D. M., Budamagunta, M. S., Voss, J. C. and Weisgraber, K. H. (2005) Modulation of apolipoprotein E structure by domain interaction: differences in lipid-bound and lipid-free forms. *J. Biol. Chem.* **280**, 34288–34295
- 88 Wolozin, B., Manger, J., Bryant, R., Cordy, J., Green, R. C. and McKee, A. (2006) Re-assessing the relationship between cholesterol, statins and Alzheimer's disease. *Acta Neurol. Scand. Suppl.* **185**, 63–70
- 89 Abildayeva, K., Jansen, P. J., Hirsch-Reinshagen, V., Bloks, V. W., Bakker, A. H., Ramaekers, F. C., de Vente, J., Groen, A. K., Wellington, C. L., Kuipers, F. and Mulder, M. (2006) 24(S)-hydroxycholesterol participates in a liver X receptor-controlled pathway in astrocytes that regulates apolipoprotein E-mediated cholesterol efflux. *J. Biol. Chem.* **281**, 12799–12808
- 90 Burgess, B. L., Parkinson, P. F., Racke, M. M., Hirsch-Reinshagen, V., Fan, J., Wong, C., Stukas, S., Theroux, L., Chan, J. Y., Donkin, J. et al. (2008) ABCG1 influences the brain cholesterol biosynthetic pathway but does not affect amyloid precursor protein or apolipoprotein E metabolism *in vivo*. *J. Lipid Res.* **49**, 1254–1267
- 91 Fan, J., Donkin, J. and Wellington, C. (2009) Greasing the wheels of $A\beta$ clearance in Alzheimer's disease: the role of lipids and apolipoprotein E. *Biofactors* **35**, 239–248
- 92 Jiang, Q., Lee, C. Y., Mandrekar, S., Wilkinson, B., Cramer, P., Zelcer, N., Mann, K., Lamb, B., Willson, T. M., Collins, J. L. et al. (2008) ApoE promotes the proteolytic degradation of $A\beta$. *Neuron* **58**, 681–693
- 93 Kim, W. S., Chan, S. L., Hill, A. F., Guillemin, G. J. and Garner, B. (2009) Impact of 27-hydroxycholesterol on amyloid- β peptide production and ATP-binding cassette transporter expression in primary human neurons. *J. Alzheimers Dis.* **16**, 121–131
- 94 Vaya, J. and Schipper, H. M. (2007) Oxysterols, cholesterol homeostasis, and Alzheimer disease. *J. Neurochem.* **102**, 1727–1737
- 95 Bell, R. D., Sagare, A. P., Friedman, A. E., Bedi, G. S., Holtzman, D. M., Deane, R. and Zlokovic, B. V. (2007) Transport pathways for clearance of human Alzheimer's amyloid β -peptide and apolipoproteins E and J in the mouse central nervous system. *J. Cereb. Blood Flow Metab.* **27**, 909–918
- 96 Deane, R., Sagare, A., Hamm, K., Parisi, M., Lane, S., Finn, M. B., Holtzman, D. M. and Zlokovic, B. V. (2008) apoE isoform-specific disruption of amyloid β peptide clearance from mouse brain. *J. Clin. Invest.* **118**, 4002–4013
- 97 Hirsch-Reinshagen, V., Chan, J. Y., Wilkinson, A., Tanaka, T., Fan, J., Ou, G., Maia, L. F., Singaraja, R. R., Hayden, M. R. and Wellington, C. L. (2007) Physiologically regulated transgenic ABCA1 does not reduce amyloid burden or amyloid- β peptide levels *in vivo*. *J. Lipid Res.* **48**, 914–923
- 98 Wahrle, S. E., Jiang, H., Parsadanian, M., Kim, J., Li, A., Knoten, A., Jain, S., Hirsch-Reinshagen, V., Wellington, C. L., Bales, K. R. et al. (2008) Overexpression of ABCA1 reduces amyloid deposition in the PDAPP mouse model of Alzheimer disease. *J. Clin. Invest.* **118**, 671–682
- 99 Eckert, G. P., Vardanian, L., Rebeck, G. W. and Burns, M. P. (2007) Regulation of central nervous system cholesterol homeostasis by the liver X receptor agonist TO-901317. *Neurosci. Lett.* **423**, 47–52
- 100 Lefterov, I., Bookout, A., Wang, Z., Staufenbiel, M., Mangelsdorf, D. and Koldamova, R. (2007) Expression profiling in APP23 mouse brain: inhibition of $A\beta$ amyloidosis and inflammation in response to LXR agonist treatment. *Mol. Neurodegener.* **2**, 20
- 101 Riddell, D. R., Zhou, H., Comery, T. A., Kouranova, E., Lo, C. F., Warwick, H. K., Ring, R. H., Kirksey, Y., Aschmies, S., Xu, J. et al. (2007) The LXR agonist TO901317 selectively lowers hippocampal $A\beta$ 42 and improves memory in the Tg2576 mouse model of Alzheimer's disease. *Mol. Cell. Neurosci.* **34**, 621–628
- 102 Mahley, R. W., Huang, Y. and Weisgraber, K. H. (2006) Putting cholesterol in its place: apoE and reverse cholesterol transport. *J. Clin. Invest.* **116**, 1226–1229
- 103 Gofman, J. W. (1950) Blood lipoproteins and atherosclerosis. *J. Clin. Invest.* **29**, 815–816
- 104 Gofman, J. W., Jones, H. B., Lindgren, F. T., Lyon, T. P., Elliott, H. A. and Strisower, B. (1950) Blood lipids and human atherosclerosis. *Circulation* **2**, 161–178
- 105 Gofman, J. W. and Lindgren, F. (1950) The role of lipids and lipoproteins in atherosclerosis. *Science* **111**, 166–171
- 106 Rader, D. J., Cohen, J. and Hobbs, H. H. (2003) Monogenic hypercholesterolemia: new insights in pathogenesis and treatment. *J. Clin. Invest.* **111**, 1795–1803
- 107 Goldberg, I. J., Kako, Y. and Lutz, E. P. (2000) Responses to eating: lipoproteins, lipolytic products and atherosclerosis. *Curr. Opin. Lipidol.* **11**, 235–241
- 108 Goldberg, I. J. and Merkel, M. (2001) Lipoprotein lipase: physiology, biochemistry, and molecular biology. *Front. Biosci.* **6**, D388–D405
- 109 Lopez-Miranda, J., Perez-Martinez, P., Marin, C., Moreno, J. A., Gomez, P. and Perez-Jimenez, F. (2006) Postprandial lipoprotein metabolism, genes and risk of cardiovascular disease. *Curr. Opin. Lipidol.* **17**, 132–138
- 110 Merkel, M., Eckel, R. H. and Goldberg, I. J. (2002) Lipoprotein lipase: genetics, lipid uptake, and regulation. *J. Lipid Res.* **43**, 1997–2006
- 111 Otarod, J. K. and Goldberg, I. J. (2004) Lipoprotein lipase and its role in regulation of plasma lipoproteins and cardiac risk. *Curr. Atheroscler. Rep.* **6**, 335–342
- 112 de la Torre, J. C. (2006) How do heart disease and stroke become risk factors for Alzheimer's disease? *Neurol. Res.* **28**, 637–644
- 113 Decarli, C. (2004) Vascular factors in dementia: an overview. *J. Neurol. Sci.* **226**, 19–23
- 114 Mielke, M. M. and Lyketsos, C. G. (2006) Lipids and the pathogenesis of Alzheimer's disease: is there a link? *Int. Rev. Psychiatry* **18**, 173–186
- 115 Panza, F., D'Introno, A., Colacicco, A. M., Capurso, C., Pichichero, G., Capurso, S. A., Capurso, A. and Solfrizzi, V. (2006) Lipid metabolism in cognitive decline and dementia. *Brain Res. Brain Res. Rev.* **51**, 275–292
- 116 Sjogren, M., Mielke, M., Gustafson, D., Zandi, P. and Skoog, I. (2006) Cholesterol and Alzheimer's disease—is there a relation? *Mech. Ageing Dev.* **127**, 138–147
- 117 Skoog, I. and Gustafson, D. (2006) Update on hypertension and Alzheimer's disease. *Neurol. Res.* **28**, 605–611
- 118 Wolozin, B. and Bednar, M. M. (2006) Interventions for heart disease and their effects on Alzheimer's disease. *Neurol. Res.* **28**, 630–636
- 119 Hartmann, T., Kuchenbecker, J. and Grimm, M. O. (2007) Alzheimer's disease: the lipid connection. *J. Neurochem.* **103** (Suppl. 1), 159–170
- 120 Hirsch-Reinshagen, V., Burgess, B. L. and Wellington, C. L. (2009) Why lipids are important for Alzheimer disease? *Mol. Cell. Biochem.* **326**, 121–129
- 121 Kivipelto, M. and Solomon, A. (2006) Cholesterol as a risk factor for Alzheimer's disease: epidemiological evidence. *Acta Neurol. Scand. Suppl.* **185**, 50–57
- 122 Lesser, G. T., Haroutunian, V., Purohit, D. P., Schneider Beerli, M., Schmeidler, J., Honkanen, L., Neufeld, R. and Libow, L. S. (2009) Serum lipids are related to Alzheimer's pathology in nursing home residents. *Dement. Geriatr. Cogn. Disord.* **27**, 42–49
- 123 Liu, J. P., Tang, Y., Zhou, S., Toh, B. H., McLean, C. and Li, H. (2010) Cholesterol involvement in the pathogenesis of neurodegenerative diseases. *Mol. Cell. Neurosci.* **43**, 33–42
- 124 Panza, F., Capurso, C., D'Introno, A., Colacicco, A. M., Vasquez, F., Pistoia, G., Capurso, A. and Solfrizzi, V. (2006) Serum total cholesterol as a biomarker for Alzheimer's disease: mid-life or late-life determinations? *Exp. Gerontol.* **41**, 805–806

- 125 Kandiah, N. and Feldman, H. H. (2009) Therapeutic potential of statins in Alzheimer's disease. *J. Neurol. Sci.* **283**, 230–234
- 126 Purandare, N. (2009) Preventing dementia: role of vascular risk factors and cerebral emboli. *Br. Med. Bull.* **91**, 49–59
- 127 Solomon, A. and Kivipelto, M. (2009) Cholesterol-modifying strategies for Alzheimer's disease. *Expert Rev. Neurotherapeutics* **9**, 695–709
- 128 Cankurtaran, M., Yavuz, B. B., Halil, M., Dagli, N., Cankurtaran, E. S. and Ariogul, S. (2005) Are serum lipid and lipoprotein levels related to dementia? *Arch. Gerontol. Geriatr.* **41**, 31–39
- 129 Raffaitin, C., Gin, H., Empana, J. P., Helmer, C., Berr, C., Tzourio, C., Portet, F., Dartigues, J. F., Alperovitch, A. and Barberger-Gateau, P. (2009) Metabolic syndrome and risk for incident Alzheimer's disease or vascular dementia: the Three-City Study. *Diabetes Care* **32**, 169–174
- 130 Sabbagh, M., Zahiri, H. R., Ceimo, J., Cooper, K., Gaul, W., Connor, D. and Sparks, D. L. (2004) Is there a characteristic lipid profile in Alzheimer's disease? *J. Alzheimers Dis.* **6**, 585–589
- 131 Razay, G., Vreugdenhil, A. and Wilcock, G. (2007) The metabolic syndrome and Alzheimer disease. *Arch. Neurol.* **64**, 93–96
- 132 Nguyen, H. N., Son, D. J., Lee, J. W., Hwang, D. Y., Kim, Y. K., Cho, J. S., Lee, U. S., Yoo, H. S., Moon, D. C., Oh, K. W. and Hong, J. T. (2006) Mutant presenilin 2 causes abnormality in the brain lipid profile in the development of Alzheimer's disease. *Arch. Pharm. Res.* **29**, 884–889
- 133 Burgess, B. L., McIsaac, S. A., Naus, K. E., Chan, J. Y., Tansley, G. H., Yang, J., Miao, F., Ross, C. J., van Eck, M., Hayden, M. R. et al. (2006) Elevated plasma triglyceride levels precede amyloid deposition in Alzheimer's disease mouse models with abundant A β in plasma. *Neurobiol. Dis.* **24**, 114–127
- 134 Liu, Y., Yang, L., Conde-Knape, K., Behr, D., Shearman, M. S. and Shachter, N. S. (2004) Fatty acids increase presenilin-1 levels and γ -secretase activity in PSwt-1 cells. *J. Lipid Res.* **45**, 2368–2376
- 135 Patil, S. and Chan, C. (2005) Palmitic and stearic fatty acids induce Alzheimer-like hyperphosphorylation of tau in primary rat cortical neurons. *Neurosci. Lett.* **384**, 288–293
- 136 Panza, F., Capurso, C., D'Introno, A., Colacicco, A. M., Del Parigi, A., Seripa, D., Pilotto, A., Capurso, A. and Solfrizzi, V. (2006) Diet, cholesterol metabolism, and Alzheimer's disease: apolipoprotein E as a possible link? *J. Am. Geriatr. Soc.* **54**, 1963–1965
- 137 Phivilay, A., Julien, C., Tremblay, C., Berthiaume, L., Julien, P., Giguere, Y. and Calon, F. (2009) High dietary consumption of trans fatty acids decreases brain docosahexaenoic acid but does not alter amyloid- β and tau pathologies in the 3xTg-AD model of Alzheimer's disease. *Neuroscience* **159**, 296–307
- 138 Florent-Bechard, S., Desbene, C., Garcia, P., Allouche, A., Youssef, I., Escanye, M. C., Koziel, V., Hanse, M., Malaplate-Armand, C., Stenger, C. et al. (2009) The essential role of lipids in Alzheimer's disease. *Biochimie* **91**, 804–809
- 139 Fotuhi, M., Mohassel, P. and Yaffe, K. (2009) Fish consumption, long-chain omega-3 fatty acids and risk of cognitive decline or Alzheimer disease: a complex association. *Nat. Clin. Pract. Neurol.* **5**, 140–152
- 140 Hooijmans, C. R. and Kiliaan, A. J. (2008) Fatty acids, lipid metabolism and Alzheimer pathology. *Eur. J. Pharmacol.* **585**, 176–196
- 141 Pauwels, E. K., Volterrani, D., Mariani, G. and Kairemo, K. (2009) Fatty acid facts, Part IV: docosahexaenoic acid and Alzheimer's disease. A story of mice, men and fish. *Drug* **22**, 205–213
- 142 Boudraut, C., Bazinet, R. P. and Ma, D. W. (2009) Experimental models and mechanisms underlying the protective effects of n-3 polyunsaturated fatty acids in Alzheimer's disease. *J. Nutr. Biochem.* **20**, 1–10
- 143 Das, U. N. (2008) Folic acid and polyunsaturated fatty acids improve cognitive function and prevent depression, dementia, and Alzheimer's disease: but how and why? *Prostaglandins Leukotrienes Essent. Fatty Acids* **78**, 11–19
- 144 Wang, L., Gill, R., Pedersen, T. L., Higgins, L. J., Newman, J. W. and Rutledge, J. C. (2009) Triglyceride-rich lipoprotein lipolysis releases neutral and oxidized FFAs that induce endothelial cell inflammation. *J. Lipid Res.* **50**, 204–213
- 145 Katz, R., Hamilton, J. A., Pownall, H. J., Deckelbaum, R. J., Hillard, C. J., Leboeuf, R. C. and Watkins, P. A. (2007) Brain uptake and utilization of fatty acids, lipids and lipoproteins: recommendations for future research. *J. Mol. Neurosci.* **33**, 146–150
- 146 Bourre, J. M. (2006) Effects of nutrients (in food) on the structure and function of the nervous system: update on dietary requirements for brain. Part 2: macronutrients. *J. Nutr. Health Aging* **10**, 386–399
- 147 Chahoud, G., Aude, Y. W. and Mehta, J. L. (2004) Dietary recommendations in the prevention and treatment of coronary heart disease: do we have the ideal diet yet? *Am. J. Cardiol.* **94**, 1260–1267
- 148 Heininger, K. (2000) A unifying hypothesis of Alzheimer's disease. III. Risk factors. *Hum. Psychopharmacol.* **15**, 1–70
- 149 Kawas, C. H. (2006) Diet and the risk for Alzheimer's disease. *Ann. Neurol.* **59**, 877–879
- 150 Morris, M. C., Evans, D. A., Bienias, J. L., Tangney, C. C., Bennett, D. A., Wilson, R. S., Aggarwal, N. and Schneider, J. (2003) Consumption of fish and n-3 fatty acids and risk of incident Alzheimer disease. *Arch. Neurol.* **60**, 940–946
- 151 Panza, F., Capurso, C. and Solfrizzi, V. (2006) Cardiovascular factors and cognitive impairment: a role for unsaturated fatty acids and Mediterranean diet? *Am. J. Cardiol.* **98**, 1120–1121
- 152 Scarneas, N., Stern, Y., Tang, M. X., Mayeux, R. and Luchsinger, J. A. (2006) Mediterranean diet and risk for Alzheimer's disease. *Ann. Neurol.* **59**, 912–921
- 153 Serra-Majem, L., Roman, B. and Estruch, R. (2006) Scientific evidence of interventions using the Mediterranean diet: a systematic review. *Nutr. Rev.* **64**, S27–S47
- 154 Solfrizzi, V., Capurso, C. and Panza, F. (2006) Adherence to a Mediterranean dietary pattern and risk of Alzheimer's disease. *Ann. Neurol.* **60**, 620
- 155 Staehelin, H. B. (2005) Micronutrients and Alzheimer's disease. *Proc. Nutr. Soc.* **64**, 565–570
- 156 Weisburger, J. H. (2002) Lifestyle, health and disease prevention: the underlying mechanisms. *Eur. J. Cancer Prev.* **11** (Suppl. 2), S1–S7
- 157 Willett, W. C. (2006) The Mediterranean diet: science and practice. *Public Health Nutr.* **9**, 105–110
- 158 Anekonda, T. S. (2006) Resveratrol—a boon for treating Alzheimer's disease? *Brain Res. Brain Res. Rev.* **52**, 316–326
- 159 Panza, F., Capurso, C., D'Introno, A., Colacicco, A. M., Parigi, A. D., Gagliardi, G., Breglia, G., Capurso, A. and Solfrizzi, V. (2007) Mediterranean diet, mild cognitive impairment, and Alzheimer's disease. *Exp. Gerontol.* **42**, 6–7
- 160 Panza, F., Solfrizzi, V., Colacicco, A. M., D'Introno, A., Capurso, C., Torres, F., Del Parigi, A., Capurso, S. and Capurso, A. (2004) Mediterranean diet and cognitive decline. *Public Health Nutr.* **7**, 959–963
- 161 Petot, G. J. and Friedland, R. P. (2004) Lipids, diet and Alzheimer disease: an extended summary. *J. Neurol. Sci.* **226**, 31–33
- 162 Solfrizzi, V., Panza, F. and Capurso, A. (2003) The role of diet in cognitive decline. *J. Neural Transm.* **110**, 95–110
- 163 Steele, M., Stuchbury, G. and Munch, G. (2007) The molecular basis of the prevention of Alzheimer's disease through healthy nutrition. *Exp. Gerontol.* **42**, 28–36
- 164 Veurink, G., Fuller, S. J., Atwood, C. S. and Martins, R. N. (2003) Genetics, lifestyle and the roles of amyloid β and oxidative stress in Alzheimer's disease. *Ann. Hum. Biol.* **30**, 639–667
- 165 Schaefer, E. J., Bongard, V., Beiser, A. S., Lamon-Fava, S., Robins, S. J., Au, R., Tucker, K. L., Kyle, D. J., Wilson, P. W. and Wolf, P. A. (2006) Plasma phosphatidylcholine docosahexaenoic acid content and risk of dementia and Alzheimer disease: the framingham heart study. *Arch. Neurol.* **63**, 1545–1550

- 166 Eiselein, L., Wilson, D. W., Lame, M. W. and Rutledge, J. C. (2007) Lipolysis products from triglyceride-rich lipoproteins increase endothelial permeability, perturb zonula occludens-1 and F-actin, and induce apoptosis. *Am. J. Physiol. Heart Circ. Physiol.* **292**, H2745–H2753
- 167 Rutledge, J. C., Mullick, A. E., Gardner, G. and Goldberg, I. J. (2000) Direct visualization of lipid deposition and reverse lipid transport in a perfused artery: roles of VLDL and HDL. *Circ. Res.* **86**, 768–773
- 168 Rutledge, J. C., Woo, M. M., Rezaei, A. A., Curtiss, L. K. and Goldberg, I. J. (1997) Lipoprotein lipase increases lipoprotein binding to the artery wall and increases endothelial layer permeability by formation of lipolysis products. *Circ. Res.* **80**, 819–828
- 169 Saraswathi, V. and Hastay, A. H. (2006) The role of lipolysis in mediating the proinflammatory effects of very low density lipoproteins in mouse peritoneal macrophages. *J. Lipid Res.* **47**, 1406–1415
- 170 Ferrante, A., Robinson, B. S., Singh, H., Jersmann, H. P., Ferrante, J. V., Huang, Z. H., Trout, N. A., Pitt, M. J., Rathjen, D. A., Easton, C. J. et al. (2006) A novel β -oxa polyunsaturated fatty acid downregulates the activation of the I κ B kinase/nuclear factor κ B pathway, inhibits expression of endothelial cell adhesion molecules, and depresses inflammation. *Circ. Res.* **99**, 34–41
- 171 Norata, G. D., Grigore, L., Raselli, S., Redaelli, L., Hamsten, A., Maggi, F., Eriksson, P. and Catapano, A. L. (2007) Post-prandial endothelial dysfunction in hypertriglyceridemic subjects: molecular mechanisms and gene expression studies. *Atherosclerosis* **193**, 321–327
- 172 Norata, G. D., Grigore, L., Raselli, S., Seccomandi, P. M., Hamsten, A., Maggi, F. M., Eriksson, P. and Catapano, A. L. (2006) Triglyceride-rich lipoproteins from hypertriglyceridemic subjects induce a pro-inflammatory response in the endothelium: molecular mechanisms and gene expression studies. *J. Mol. Cell. Cardiol.* **40**, 484–494
- 173 Norata, G. D., Marchesi, P., Passamonti, S., Pirillo, A., Violi, F. and Catapano, A. L. (2007) Anti-inflammatory and anti-atherogenic effects of catechin, caffeic acid and trans-resveratrol in apolipoprotein E deficient mice. *Atherosclerosis* **191**, 265–271
- 174 Ziouzenkova, O., Asatryan, L., Sahady, D., Orasanu, G., Perrey, S., Cutak, B., Hassell, T., Akiyama, T. E., Berger, J. P., Sevanian, A. and Plutzky, J. (2003) Dual roles for lipolysis and oxidation in peroxisome proliferation-activator responses to electronegative low density lipoprotein. *J. Biol. Chem.* **278**, 39874–39881
- 175 Ziouzenkova, O., Perrey, S., Asatryan, L., Hwang, J., MacNaul, K. L., Moller, D. E., Rader, D. J., Sevanian, A., Zechner, R., Hoefler, G. and Plutzky, J. (2003) Lipolysis of triglyceride-rich lipoproteins generates PPAR ligands: evidence for an antiinflammatory role for lipoprotein lipase. *Proc. Natl. Acad. Sci. U.S.A.* **100**, 2730–2735
- 176 Lee, J. and Goldberg, I. J. (2007) Lipoprotein lipase-derived fatty acids: physiology and dysfunction. *Curr. Hypertens. Rep.* **9**, 462–466
- 177 Takahashi, M., Hiyama, Y., Yokoyama, M., Yu, S., Hu, Y., Melford, K., Bensadoun, A. and Goldberg, I. J. (2008) *In vivo* arterial lipoprotein lipase expression augments inflammatory responses and impairs vascular dilatation. *Arterioscler. Thromb. Vasc. Biol.* **28**, 455–462
- 178 Esenabhalu, V. E., Cerimagic, M., Malli, R., Osibow, K., Levak-Frank, S., Frieden, M., Sattler, W., Kostner, G. M., Zechner, R. and Graier, W. F. (2002) Tissue-specific expression of human lipoprotein lipase in the vascular system affects vascular reactivity in transgenic mice. *Br. J. Pharmacol.* **135**, 143–154
- 179 Borradaile, N. M., Han, X., Harp, J. D., Gale, S. E., Ory, D. S. and Schaffer, J. E. (2006) Disruption of endoplasmic reticulum structure and integrity in lipotoxic cell death. *J. Lipid Res.* **47**, 2726–2737
- 180 Cnop, M., Ladriere, L., Hekerman, P., Ortis, F., Cardozo, A. K., Dogusan, Z., Flamez, D., Boyce, M., Yuan, J. and Eizirik, D. L. (2007) Selective inhibition of eukaryotic translation initiation factor 2 α dephosphorylation potentiates fatty acid-induced endoplasmic reticulum stress and causes pancreatic β -cell dysfunction and apoptosis. *J. Biol. Chem.* **282**, 3989–3997
- 181 Wei, Y., Wang, D. and Pagliassotti, M. J. (2007) Saturated fatty acid-mediated endoplasmic reticulum stress and apoptosis are augmented by *trans*-10, *cis*-12-conjugated linoleic acid in liver cells. *Mol. Cell. Biochem.* **303**, 105–113
- 182 Wei, Y., Wang, D., Topczewski, F. and Pagliassotti, M. J. (2006) Saturated fatty acids induce endoplasmic reticulum stress and apoptosis independently of ceramide in liver cells. *Am. J. Physiol. Endocrinol. Metab.* **291**, E275–E281
- 183 Borradaile, N. M., Buhman, K. K., Listenberger, L. L., Magee, C. J., Morimoto, E. T., Ory, D. S. and Schaffer, J. E. (2006) A critical role for eukaryotic elongation factor 1A-1 in lipotoxic cell death. *Mol. Biol. Cell* **17**, 770–778
- 184 Wang, H., Kouri, G. and Wollheim, C. B. (2005) ER stress and SREBP-1 activation are implicated in β -cell glucolipotoxicity. *J. Cell Sci.* **118**, 3905–3915
- 185 Hoeks, J., Hesselink, M. K. and Schrauwen, P. (2006) Involvement of UCP3 in mild uncoupling and lipotoxicity. *Exp. Gerontol.* **41**, 658–662
- 186 Lionetti, L., Mollica, M. P., Crescenzo, R., D'Andrea, E., Ferraro, M., Bianco, F., Liverini, G. and Iossa, S. (2007) Skeletal muscle subsarcolemmal mitochondrial dysfunction in high-fat fed rats exhibiting impaired glucose homeostasis. *Int. J. Obes.* **31**, 1596–1604
- 187 Schrauwen, P. (2007) High-fat diet, muscular lipotoxicity and insulin resistance. *Proc. Nutr. Soc.* **66**, 33–41
- 188 Summers, S. A. (2006) Ceramides in insulin resistance and lipotoxicity. *Prog. Lipid Res.* **45**, 42–72
- 189 Feldstein, A. E., Werneburg, N. W., Canbay, A., Guicciardi, M. E., Bronk, S. F., Rydzewski, R., Burgart, L. J. and Gores, G. J. (2004) Free fatty acids promote hepatic lipotoxicity by stimulating TNF- α expression via a lysosomal pathway. *Hepatology* **40**, 185–194
- 190 Feldstein, A. E., Werneburg, N. W., Li, Z., Bronk, S. F. and Gores, G. J. (2006) Bax inhibition protects against free fatty acid-induced lysosomal permeabilization. *Am. J. Physiol. Gastrointest. Liver Physiol.* **290**, G1339–G1346
- 191 Emerit, J., Edeas, M. and Bricaire, F. (2004) Neurodegenerative diseases and oxidative stress. *Biomed. Pharmacother.* **58**, 39–46
- 192 Floyd, R. A. (1999) Antioxidants, oxidative stress, and degenerative neurological disorders. *Proc. Soc. Exp. Biol. Med.* **222**, 236–245
- 193 Picklo, Sr, M. J. and Montine, T. J. (2007) Mitochondrial effects of lipid-derived neurotoxins. *J. Alzheimers Dis.* **12**, 185–193
- 194 Laskowitz, D. T., Fillit, H., Yeung, N., Toku, K. and Vitek, M. P. (2006) Apolipoprotein E-derived peptides reduce CNS inflammation: implications for therapy of neurological disease. *Acta Neurol. Scand. Suppl.* **185**, 15–20
- 195 Patil, S., Sheng, L., Masserang, A. and Chan, C. (2006) Palmitic acid-treated astrocytes induce BACE1 upregulation and accumulation of C-terminal fragment of APP in primary cortical neurons. *Neurosci. Lett.* **406**, 55–59
- 196 Calabrese, V., Guagliano, E., Sapienza, M., Panebianco, M., Calafato, S., Puleo, E., Pennisi, G., Mancuso, C., Butterfield, D. A. and Stella, A. G. (2007) Redox regulation of cellular stress response in aging and neurodegenerative disorders: role of vitagenes. *Neurochem. Res.* **32**, 757–773
- 197 Kim, H. S., Kong, K. A., Chung, H., Park, S. and Kim, M. H. (2007) ER stress induces the expression of Jpk, which inhibits cell cycle progression in F9 teratocarcinoma cell. *Ann. N.Y. Acad. Sci.* **1095**, 76–81
- 198 Schippling, S., Kontush, A., Arlt, S., Buhmann, C., Sturenburg, H. J., Mann, U., Muller-Thomsen, T. and Beisiegel, U. (2000) Increased lipoprotein oxidation in Alzheimer's disease. *Free Radical Biol. Med.* **28**, 351–360
- 199 Wang, J. Y., Wen, L. L., Huang, Y. N., Chen, Y. T. and Ku, M. C. (2006) Dual effects of antioxidants in neurodegeneration: direct neuroprotection against oxidative stress and indirect protection via suppression of glia-mediated inflammation. *Curr. Pharmaceut. Design* **12**, 3521–3533

- 200 Hong, H., Zeng, J. S., Kreulen, D. L., Kaufman, D. I. and Chen, A. F. (2006) Atorvastatin protects against cerebral infarction via inhibition of NADPH oxidase-derived superoxide in ischemic stroke. *Am. J. Physiol. Heart Circ. Physiol.* **291**, H2210–H2215
- 201 Wang, X., Chen, S., Ma, G., Ye, M. and Lu, G. (2005) Involvement of proinflammatory factors, apoptosis, caspase-3 activation and Ca^{2+} disturbance in microglia activation-mediated dopaminergic cell degeneration. *Mech. Ageing Dev.* **126**, 1241–1254
- 202 Ouyang, Y. B. and Giffard, R. G. (2004) Changes in astrocyte mitochondrial function with stress: effects of Bcl-2 family proteins. *Neurochem. Int.* **45**, 371–379
- 203 Wang, L., Sapuri-Butti, A. R., Aung, H. H., Parikh, A. N. and Rutledge, J. C. (2008) Triglyceride-rich lipoprotein lipolysis increases aggregation of endothelial cell membrane microdomains and produces reactive oxygen species. *Am. J. Physiol. Heart Circ. Physiol.* **295**, H237–H244
- 204 Jin, J. J., Kim, H. D., Maxwell, J. A., Li, L. and Fukuchi, K. (2008) Toll-like receptor 4-dependent upregulation of cytokines in a transgenic mouse model of Alzheimer's disease. *J. Neuroinflammation* **5**, 23
- 205 van Noort, J. M. and Bsibsi, M. (2009) Toll-like receptors in the CNS: implications for neurodegeneration and repair. *Progr. Brain Res.* **175**, 139–148
- 206 Wong, S. W., Kwon, M. J., Choi, A. M., Kim, H. P., Nakahira, K. and Hwang, D. H. (2009) Fatty acids modulate Toll-like receptor 4 activation through regulation of receptor dimerization and recruitment into lipid rafts in a reactive oxygen species-dependent manner. *J. Biol. Chem.* **284**, 27384–27392
- 207 Adibhatla, R. M. and Hatcher, J. F. (2008) Altered lipid metabolism in brain injury and disorders. *Subcell. Biochem.* **49**, 241–268
- 208 Bazan, N. G. (2007) Omega-3 fatty acids, pro-inflammatory signaling and neuroprotection. *Curr. Opin. Clin. Nutr. Metab. Care* **10**, 136–141
- 209 Bieschke, J., Zhang, Q., Bosco, D. A., Lerner, R. A., Powers, E. T., Wentworth, Jr, P. and Kelly, J. W. (2006) Small molecule oxidation products trigger disease-associated protein misfolding. *Accounts Chem. Res.* **39**, 611–619
- 210 Farooqui, A. A., Horrocks, L. A. and Farooqui, T. (2007) Interactions between neural membrane glycerophospholipid and sphingolipid mediators: a recipe for neural cell survival or suicide. *J. Neurosci. Res.* **85**, 1834–1850
- 211 Farooqui, A. A., Ong, W. Y., Horrocks, L. A., Chen, P. and Farooqui, T. (2007) Comparison of biochemical effects of statins and fish oil in brain: the battle of the titans. *Brain Res. Rev.* **56**, 443–471
- 212 Jaeger, S. and Pietrzik, C. U. (2008) Functional role of lipoprotein receptors in Alzheimer's disease. *Curr. Alzheimer Res.* **5**, 15–25
- 213 Kapoor, M., Shaw, O. and Appleton, I. (2005) Possible anti-inflammatory role of COX-2-derived prostaglandins: implications for inflammation research. *Curr. Opin. Investig. Drugs.* **6**, 461–466
- 214 Leitinger, N. (2008) The role of phospholipid oxidation products in inflammatory and autoimmune diseases: evidence from animal models and in humans. *Subcell. Biochem.* **49**, 325–350
- 215 Lukiw, W. J. and Bazan, N. G. (2008) Docosahexaenoic acid and the aging brain. *J. Nutr.* **138**, 2510–2514
- 216 Mocchetti, I. and Brown, M. (2008) Targeting neurotrophin receptors in the central nervous system. *CNS Neurol. Disorders Drug Targets* **7**, 71–82
- 217 Mukherjee, P. K., Chawla, A., Loayza, M. S. and Bazan, N. G. (2007) Docosanoids are multifunctional regulators of neural cell integrity and fate: significance in aging and disease. *Prostaglandins Leukotrienes Essent. Fatty Acids* **77**, 233–238
- 218 Pomponi, M., Bria, P. and Pomponi, M. (2008) Is Alzheimer's disease a synaptic disorder? *J. Alzheimers Dis.* **13**, 39–47
- 219 Montine, T. J., Quinn, J., Kaye, J. and Morrow, J. D. (2007) F_2 -isoprostanes as biomarkers of late-onset Alzheimer's disease. *J. Mol. Neurosci.* **33**, 114–119
- 220 Farooqui, A. A. (2009) Lipid mediators in the neural cell nucleus: their metabolism, signaling, and association with neurological disorders. *Neuroscientist* **15**, 392–407
- 221 Serhan, C. N., Hong, S., Gronert, K., Colgan, S. P., Devchand, P. R., Mirick, G. and Moussignac, R. L. (2002) Resolvins: a family of bioactive products of omega-3 fatty acid transformation circuits initiated by aspirin treatment that counter proinflammation signals. *J. Exp. Med.* **196**, 1025–1037
- 222 Tetali, S. D., Budamagunta, M. S., Simion, C., den Hartigh, L. J., Kálai, T., Hideg, K., Hatters, D. M., Weisgraber, K. H., Voss, J. C. and Rutledge, J. C. (2010) VLDL lipolysis products increase VLDL fluidity and convert apolipoprotein E4 into a more expanded conformation. *J. Lipid Res.* **51**, 1273–1283

Received 4 February 2010/2 June 2010; accepted 14 June 2010

Published on the Internet 5 August 2010, doi:10.1042/CS20100094